# Oragraft® Prime

**Clinical Overview**

OraGraft Prime is 100% bone fibres, demineralised to encourage bone formation and healing. The fibres interlock, allowing the graft to become mouldable upon rehydration without the use of a carrier.

**Applications**

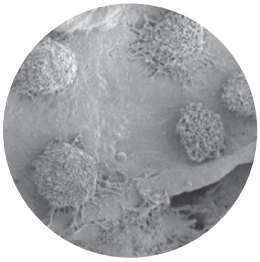
Surgical procedures that require a bone void filler

**Features & Benefits**

- **100% Bone**: Facilitates natural remodelling during the bone healing process (no human, xenograft or synthetic carriers).
- **Osteoconductive:** The large surface area and interconnected network of demineralised cortical fibres provides a scaffold that promotes cellular attachment and cell spreading.[2]
- **Osteoinductive Potential:** Optimally demineralised by LifeNet Health's patented and proprietary PAD® technology to expose natural growth factors.[3-7]
- **Versatile:** Mouldable upon rehydration to conform to the surgical site.
- **Resists Migration:** Interlocking fibres allow graft to remain intact and in place.
- **Safety:** Sterilised using proprietary and patented technology, providing a sterility assurance level of $10^{-6}$ to reduce the risk of disease transmission without compromising the graft's inherent osteoconductive properties or osteoinductive potential.[8]
- **Convenience:** Ambient storage and rapid rehydration.

*100% bone fibres*

*Mouldable upon rehydration*



*Hospitable environment for bone growth (cell attachment at one hour)*


Speak to your local Business Development Manager for further information or contact us using the details below:

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## OraGraft Prime

Freeze-dried (10˚C to 30˚C)

| Volume | Order Code | Shelf Life |
|---|---|---|
| 0.5 cc | DF-1007 | 4 years |
| 1.0 cc | DF-1008 | 4 years |
| 2.5 cc | DF-1009 | 5 years |

Instructions for use available at **LifeNetHealth.org/IFU**


**References**

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2. Murphy MB, Suzuki RK, Sand TT, et al. Short term culture of mesenchymal stem cells with commercial osteoconductive carriers provides unique insights into biocompatibility. J Clin. Med. 2013; 2,49-66; doi:10.3390/jcm2030049
3. Zhang M, Powers RM, and Wolfinbarger L. Effect(s) of the demineralization process on the osteoinductivity of demineralized bone matrix. J Periodontol. 1997; 68:1085-1092
4. Turonis JW, McPherson JC 3rd, Cuenin MF, et al. The effect of residual calcium in decalcified freeze-dried bone allograft in a critical-sized defect in the Rattus norvegicus calvarium. J Oral Implantol. 2006; 32(2):55-62
5. Herold RW, Pashley DH, Cuenin MF, et al. The effects of Varying degrees of Allograft Decalcification on Cultured Porcine Osteoclast cells. J Periodontol. 2002 Feb; 73(2):213-9
6. Mott DA, Mailhot J, Cuenin MF, et al. Enhancement of osteoblast proliferation in vitro by selective enrichment of demineralized freeze-dried bone allograft with specific growth factors. J Oral Implantol. 2002; 28(2):57-66
7. Pietrzak WS, Ali SN, Chitturi D, et al. BMP depletion occurs during prolonged acid demineralization of bone: characterization and implications for graft preparation. Cell Tiss. Bank. 2007 (Published on line)
8. Eisenlohr LM. “Allograft Tissue Sterilization Using Allowash XG (R) .” 2007 Bio-Implants Brief.



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